



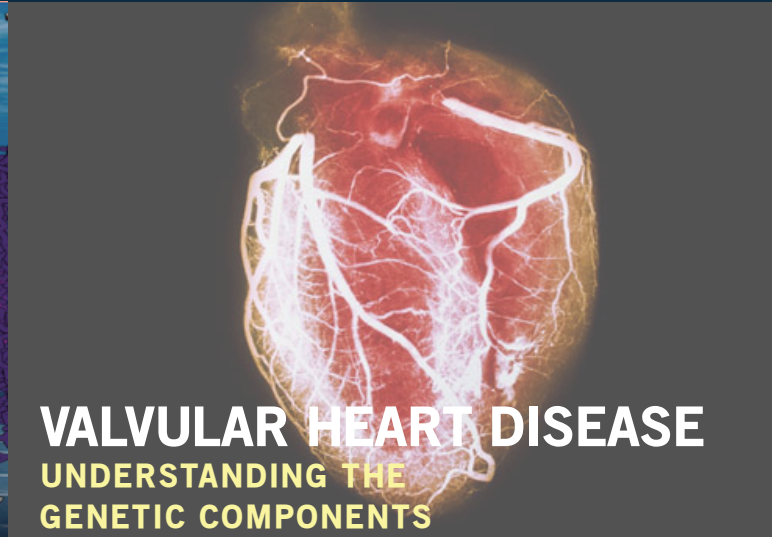
## BALOXAVIR MARBOXIL: A NOVEL ANTI-INFLUENZA DRUG

DANIEL RAYNER

Over the last couple of decades, only two classes of drugs have been approved for use in patients with influenza: M2 ion-channel inhibitors (e.g., rimantadine) and neuraminidase inhibitors (e.g., oseltamivir).<sup>1</sup> However, resistance to these drugs has been increasing, with circulating influenza strains now predominantly resistant to M2 ion-channel inhibitors.<sup>2</sup> Even resistance towards the current frontline for influenza treatment, neuraminidase inhibitors, is a health concern, as demonstrated by the oseltamivir-resistant influenza A(H1N1) pandemic during the 2008-2009 season.<sup>2,3</sup>

Approved by the U.S. Food and Drug Administration (FDA) in October 2018, baloxavir marboxil (BM) is a novel anti-influenza drug that targets the RNA-dependent RNA polymerase (RdRp), the heterotrimeric enzyme responsible for transcription of viral mRNA.<sup>2,4</sup> Specifically, BM inhibits the polymerase acidic (PA) protein, the subunit responsible for cleaving the 5' end of host pre-mRNA, which is subsequently used as a primer for viral mRNA production.<sup>5</sup> However, BM is not exempt from resistance mutations — I38T/M/F point mutations in the PA subunit were observed in 2.2% and 9.7% of BM recipients in phase II and phase III clinical trials, respectively.<sup>2</sup> These substitutions were found to reduce the viral strains' susceptibility to BM by more than a factor of ten in those infected with influenza A(H1N1).<sup>2</sup>

The presence of a new anti-influenza drug on the market may allow for the use of combination therapies for patients with complicated influenza infections.<sup>6</sup> The combination of BM and oseltamivir was more effective than each monotherapy in treating influenza A in murine models, and human combination studies are currently underway.<sup>6,7</sup>



## VALVULAR HEART DISEASE UNDERSTANDING THE GENETIC COMPONENTS

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Occurring in approximately 2.5% of the general population, valvular heart disease (VHD) is a group of cardiovascular diseases involving the damage or defect of one of the four heart valves: pulmonary, aortic, tricuspid, and mitral.<sup>1,2</sup> Despite its high frequency, therapeutic approaches for VHD are extremely limited, with surgery being the primary treatment for valve replacement or repair.<sup>3,4</sup> Presently, only a small number of genes have been identified as monogenic causes of nonsyndromic VHD.<sup>5-7</sup>

Recently, research by Wünnemann and colleagues has led to the discovery of another monogenic VHD gene, *ADAMTS19*, which encodes an enzyme responsible for extracellular matrix modelling activity.<sup>8,9</sup> Using whole-exome sequencing on two consanguineous families with prevalent early-onset VHD, researchers found that affected individuals had homozygous, loss-of-function alleles in *ADAMTS19*.<sup>8</sup> Supplemented with an *Adamts19* knockout murine model, Wünnemann and colleagues hypothesize that loss of *Adamts19* interferes with shear stress signalling in the endothelial cells of the aortic valve, inducing upregulation of the transcription factor Klf2.<sup>8</sup> Klf2 regulates Wnt9b, the ligand responsible for the remodeling of cardiac cushions into mature heart valves.<sup>10</sup> This dysregulation of Klf2 leads to VHD through extracellular matrix disorganization, as well as increased cellularity and proteoglycan deposition in the valves.<sup>8</sup>

Improving our understanding of the genetic components, molecular pathways, and cellular mediators involved in the development of the disease may aid in treating VHD. Not only can it improve genetic screening for high-risk individuals, but it also opens up an avenue for potential VHD pharmacological therapies, which can delay or halt disease progression.<sup>11</sup>

- Mushtaq A. Baloxavir: Game-changer or much ado about nothing? *Lancet Respir Med*. 2018;6(12):903-4. Available from: doi:10.1016/S2213-2600(18)30469-7.
- Hayden FG, Sugaya N, Hirotsu N, Lee N, de Jong MD, Hurt AC, et al. Baloxavir marboxil for uncomplicated influenza in adults and adolescents. *N Engl J Med*. 2018;379(10):913-23. Available from: doi:10.1056/NEJMoa1716197.
- Lee N, Hurt A. Neuraminidase inhibitor resistance in influenza: A clinical perspective. *Curr Opin Infect Dis*. 31(6):520-6. Available from: doi:10.1097/QCO.0000000000000498.
- Yoshino R, Yasuo N, Sekijima M. Molecular dynamics simulation reveals the mechanism by which the influenza cap-dependent endonuclease acquires resistance against baloxavir marboxil. *Sci Rep*. 2019;9(1):17464. Available from: doi:10.1038/s41598-019-53945-1.
- O'Hanlon R, Shaw ML. Baloxavir marboxil: The new influenza drug on the market. *Curr Opin Virol*. 2019;35:14-8. Available from: doi:10.1016/j.coviro.2019.01.006.
- Fukao K, Noshi T, Yamamoto A, Kitano M, Ando Y, Noda T, et al. Combination treatment with the cap-dependent endonuclease inhibitor baloxavir marboxil and a neuraminidase inhibitor in a mouse model of influenza A virus infection. *J Antimicrob Chemother*. 2019;74(3):654-62. Available from: doi:10.1093/jac/dky462.
- Fujita J. Introducing the new anti-influenza drug, baloxavir marboxil. *Respir Investig*. 2020;58(1):1-3. Available from: doi:10.1016/j.resinv.2019.10.005.

- Nikomo VT, Gardin JM, Skelton TN, Gottdiener JS, Scott CG, Enriquez-Sarano M. Burden of valvular heart diseases: A population-based study. *Lancet*. 2006;368(9540):1005-11. Available from: doi:10.1016/S0140-6736(06)69208-8.
- Zeng Y, Sun R, Li X, Liu M, Chen S, Zhang P. Pathophysiology of valvular heart disease. *Exp Ther Med*. 2016;11(4):1184-8. Available from: doi:10.3892/etm.2016.3048.
- Itagaki S, Adams DH, Anyanwu AC. Triggers for surgical referral in degenerative mitral valve regurgitation. *Circ J*. 2013;77(1):28-34. Available from: doi:10.1253/circj.12-0972.
- Maeda K, Kuratani T, Mizote I, Shimamura K, Takeda Y, Torikai K, et al. Early experiences of transcatheter aortic valve replacement in Japan. *Circ J*. 2013;77(2):359-62. Available from: doi:10.1253/circj.12-0650.
- Garg V, Muth AN, Ransom JF, Schluterman MK, Barnes R, King IN, et al. Mutations in NOTCH1 cause aortic valve disease. *Nature*. 2005;437(7056):270-4. Available from: doi:10.1038/nature03940.
- Ta-Shma A, Zhang K, Salimova E, Seiro-Mosti D, Stegner D, et al. Congenital valvular defects associated with deleterious mutations in the PLD1 gene. *J Med Genet*. 2017;54(4):278-86. Available from: doi:10.1136/jmedgenet-2016-104259.
- Gould RA, Aziz H, Woods CE, Seman-Senderos MA, Sparks E, Preuss C, et al. ROBO4 variants predispose individuals to bicuspid aortic valve and thoracic aortic aneurysm. *Nat Genet*. 2019;51(1):42-50. Available from: doi:10.1038/s41588-018-0265-y.
- Wünnemann F, Ta-Shma A, Preuss C, Leclerc S, van Vliet PP, Oneglia A, et al. Loss of ADAMTS19 causes progressive non-syndromic heart valve disease. *Nat Genet*. 2020;52(1):40-7. Available from: doi:10.1038/s41588-019-0536-2.
- Brunet FG, Fraser FW, Binder MJ, Smith AD, Kintakas C, Dancevic CM, et al. The evolutionary conservation of the A disintegrin-like and metalloprotease domain with thrombospondin-1 motif metzincins across vertebrate species and their expression in teleost zebrafish. *BMC Evol Biol*. 2015;15:22. Available from: doi:10.1186/s12862-015-0281-9.
- Goddard LM, Duchemin AL, Ramalingan H, Wu B, Chen M, Barnezi S, et al. Hemodynamic forces sculpt developing heart valves through a Klf2-Wnt9b paracrine signaling axis. *Dev Cell*. 2017;43(3):274-89.e5. Available from: doi:10.1016/j.devcel.2017.09.023.
- Padang R, Bagnall RD, Semsarian C. Genetic basis of familial valvular heart disease. *Circ Cardiovasc Genet*. 2012;5(5):569-80. Available from: doi:10.1161/CIRCGENETICS.112.962894.



## THE FARNESOID X RECEPTOR A NOVEL TARGET FOR HEPATOCELLULAR CARCINOMA

## RHO KINASE INHIBITORS: MANIPULATING CELL SHAPE FOR TREATMENT OF GLAUCOMA

SHADI SADEGHIAN

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The Farnesoid X Receptor (FXR) is a nuclear receptor expressed in the liver that is known to maintain bile acid homeostasis.<sup>1</sup> Recently, FXR has also been implicated in hepatocellular carcinoma (HCC), acting as a tumour suppressor.<sup>1</sup> This effect is dependent on the activation of the IL-6/Jak-2/STAT-3 inflammatory pathway—a molecular cascade that drives tumour formation in many types of cancer.<sup>1,2</sup> It has also been established that the activation of STAT-3, a substrate in the inflammatory pathway, upregulates the expression of genes that further induce cell division and inhibit apoptosis.<sup>2,3</sup> Recently, a growing body of evidence has suggested that FXR deficiency causes STAT-3 activation, resulting in cell proliferation.<sup>2</sup>

Under FXR-deficient conditions (commonly observed in HCC) the STAT-3 pathway is activated by an increase in the expression of an inflammatory cytokine, IL-1 $\beta$ , which subsequently increases levels of IL-6. The reason for this upregulation is the increase in bile acid concentration resulting from FXR deficiency.<sup>4</sup> This bile acid build-up induces cholestasis and hepatic inflammation, thus recruiting various inflammatory cytokines. As IL-6 binds to its receptor, the IL-6/Jak-2/STAT-3 pathway is activated. As a result, newly formed STAT-3 homodimers are able to activate downstream genes responsible for hepatic carcinogenesis.<sup>2</sup> Various studies have demonstrated that FXR ligands cause an upregulation of FXR expression and reduce the cholestasis that initiates this carcinogenic cycle.<sup>3</sup> Cell cycle analyses have shown that FXR ligation increases the number of HepG2 (HCC) cells at the cell cycle arrest phase, and decreases the cells in the synthesis phase.<sup>3,5</sup> All of this demonstrates the ability of FXR to mitigate uncontrolled cell growth in the liver.<sup>3</sup>

Glaucoma is the second leading cause of irreversible blindness worldwide.<sup>1</sup> This condition involves the degeneration of retinal ganglion cells and is characterized by elevated intraocular pressure (IOP).<sup>1</sup> The problems arising in glaucoma originate in the anterior chamber of the eye, where aqueous humour (AH) drainage via the trabecular meshwork (TM) is impeded.<sup>1</sup> Since the damage to retinal ganglion cells in glaucoma is irreversible, the goal of treatment is to preserve remaining visual acuity by mitigating high IOP.<sup>2</sup>

It was first noticed in the 1990s that the pharmacological manipulation of the cytoskeleton of TM cells decreased aqueous humor outflow resistance, thus significantly reducing IOP.<sup>2</sup> Rho kinase inhibitors play a key role in the rigidity of the cytoskeleton of TM cells and thus have been of particular pharmacologic importance, as they influence AH drainage efficiency.<sup>2,3</sup> The GTPase Rho, when bound to guanosine triphosphate (GTP), is able to activate Rho kinase.<sup>3</sup> Rho kinase is then able to undergo a series of biological reactions that change the properties of the cytoskeleton, which dictates the cell's morphology.<sup>4</sup> By this mechanism, the protein increases the rigidity of the TM cells, affecting their mobility.<sup>3,4</sup> At the same time, Rho kinase inhibitors are able to modulate other morphological issues in TM cells including changes in the extracellular matrix and mitigate irregular contractile forces.<sup>4</sup> Overall, such changes allow for improved AH outflow, lowering IOP.<sup>4</sup> A randomized, placebo-controlled study demonstrated that administration of 0.25% of a Rho-kinase inhibitor twice per day reduced IOP by 28%, or around 6.8 mmHg.<sup>5</sup> Various other studies have had similar findings, demonstrating the promise of this practice.

1. Bowlus C. Obeticholic acid for the treatment of primary biliary cholangitis in adult patients: Clinical utility and patient selection. *Hepat Med*. 2016;2016(8):89-95. Available from: doi:10.2147/HMER.S91709.
2. Attia Y, Tawfiq R, Ali A, Elmazar M. The FXR agonist, obeticholic acid, suppresses HCC proliferation & metastasis: Role of IL-6/STAT3 signalling pathway. *Sci Rep*. 2017;(12502). Available from: doi:10.1038/s41598-017-12629-4.
3. Grohmann M, Wiede F, Dodd G, Gurzov E, Ooi G, Butt T, et al. Obesity drives STAT-1-dependent NASH and STAT-3-dependent HCC. *Cell*. 2018;175(5):1289-1306.e20. Available from: doi:10.1016/j.cell.2018.09.053.
4. Verbeke L, Mannaerts I, Schierwagen R, Govaere O, Klein S, Vander Elst I, et al. FXR agonist obeticholic acid reduces hepatic inflammation and fibrosis in a rat model of toxic cirrhosis. *Sci Rep*. 2016;6:33453. Available from: doi:10.1038/srep33453.
5. Staropoli J. Tumorigenesis and neurodegeneration: two sides of the same coin? *BioEssays*. 2008;30(8):719-727. Available from: doi:10.1002/bies.20784.

1. Weinreb R, Aung T, Medeiros F. The pathophysiology and treatment of glaucoma. *JAMA*. 2014;311(18):1901. Available from: doi:10.1001/jama.2014.3192.
2. Inoue K. Managing adverse effects of glaucoma medications. *Clin Ophthalmol*. 2014;9:903. Available from: doi:10.2147/OPTH.S44708.
3. Tanna A, Johnson M. Rho kinase inhibitors as a novel treatment for glaucoma and ocular hypertension. *Ophthalmology*. 2018;125(11):1741-56. Available from: doi:10.1016/j.ophtha.2018.04.040.
4. Moshirfar M, Parker L, Birdsong OC, Ronquillo YC, Hofstedt D, Shah TJ, et al. Use of rho kinase inhibitors in ophthalmology: A review of the literature. *Med Hypothesis Discov Innov Ophthalmol*. 2018;7(3):101-11. Available from: doi:https://www.ncbi.nlm.nih.gov/pubmed/30386798 [cited 2020 Feb 24].
5. Williams RD, Novack GD, van Haarlem T, Kocczynski C, AR-12286 Phase 2A Study Group. Ocular hypotensive effect of the Rho kinase inhibitor AR-12286 in patients with glaucoma and ocular hypertension. *Am J Ophthalmol*. 2011;152(5):834-41.e1. Available from: doi:10.1016/j.ajo.2011.04.012.