

sual acuity with no particular adverse cell proliferation or rejection when ESC-derived retinal pigment epithelial cells were transplanted into the subretinal space to treat age-related macular degeneration.³⁰ However, ESC therapy is limited by concerns regarding long-term safety and graft survival.

As an alternative, investigators have employed adult stem cell therapy (ASCT).³⁰ ASCT requires *ex vivo* manipulations that involve isolating, enriching, identifying, and growing adult stem cells before they can be used to replace any cells of the dysfunctional organs via transplantation and cell injection.³⁰ ASCT aims to allow normal, healthy cells to differentiate into functional cells in the target diseased tissues.^{30,32}

Specifically, DSCs can be integrated into ASCT to regenerate and restore ocular tissues. This is because DSCs are derived from cranial NCC and may possess similar properties to neural crest progenitor cells that give rise to many structures of the anterior segment of the eye.^{30,32} In fact, the study showed that when undifferentiated, immature human dental pulp stem cells (DPSCs) were transplanted into an animal model of limbal stem cell deficiency, it resulted in a reconstructed corneal epithelium, reduction in neovascularization, and clear cornea. This animal model involves extensive corneal damage and permanent visual impairment, and is often used to study the effects of stem cells in healing damaged tissues. This particular model and its limbal stem cell deficiency manifests as the lack of repopulation of corneal epithelium and is viable for testing the healing capacity of the DSCs. These results clearly demonstrate their capacity to replace limbal stem cells and restore the cornea.³⁰⁻³² Furthermore, the study showed that DPSCs may serve as an abundant source of retinal-like stem cells with the ability to differentiate into retinal neurons and photoreceptors.³⁰

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- Strungaru M, Dinu I, Walter M. Genotype-Phenotype Correlations in Axenfeld-Rieger Malformation and Glaucoma Patients with FOXC1 and PITX2 Mutations. *Investigative Ophthalmology & Visual Science*. 2007;48(1):228.
- Could DB, John SW. Anterior segment dysgenesis and the developmental glaucomas are complex traits. *Human molecular genetics*. 2002 May 15;11(10):1185-93.
- Reneker LW, Silversides DW, Xu L, Overbeek PA. Formation of corneal endothelium is essential for anterior segment development—a transgenic mouse model of anterior segment dysgenesis. *Development*. 2000 Feb 1;127(3):533-42.
- Williams Bohnsack B. Neural crest derivatives in ocular development: Discerning the eye of the storm. *Birth Defects Research Part C: Embryo Today: Reviews*. 2015;105(2):87-95.
- Beebe DC, Coats JM. The lens organizes the anterior segment: specification of neural crest cell differentiation in the avian eye. *Developmental biology*. 2000 Apr 15;220(2):424-31.
- Creuzet S, Vincent C, Couly G. Neural crest derivatives in ocular and periocular structures. *Int J Dev Biol*. 2005 Jan 1;49(2-3):161-71.
- Berry F. Functional interactions between FOXC1 and PITX2 underlie the sensitivity to FOXC1 gene dose in Axenfeld-Rieger syndrome and anterior segment dysgenesis. *Human Molecular Genetics*. 2006; 15(6): 905-919.
- Lines MA, Kozlowski K, Walter MA. Molecular genetics of Axenfeld-Rieger malformations. *Human molecular genetics*. 2002 May 15;11(10):1177-87.
- Hjalil TA, Semina EV. Current molecular understanding of Axenfeld-Rieger syndrome. *Expert reviews in molecular medicine*. 2005 Nov 8;7(25):1-7.
- Doucette L, Green J, Fernandez B, Johnson G, Parfrey P, Young T. A novel, non-stop mutation in FOXE3 causes an autosomal dominant form of variable anterior segment dysgenesis including Peters anomaly. *Eur J Hum Genet*. 2010;19(3):293-299.

CONCLUSION

Continued research on ASD has allowed for identification of multiple genes associated with the condition, several of which include *Pitx2*, *Foxc1*, *Tfap2b*. It has been found that these genes do not work independently of one another. Rather, they regulate or affect one another in the process of POM specification into anterior segment tissues such as the components of the corneal layers and structures of the iridocorneal angle. Research has also suggested the possibility of sequential formation of anterior segment tissues. This was clearly demonstrated in the lens ablation experiment, in which the lens allowed for subsequent specification of the POM into the anterior structures through inductive signaling. This review paper aimed to emphasize the importance of proper embryonic development of the anterior structures and the possible complications that can arise as a result of its improper development. These complications include ARS, subcapsular cataracts, and glaucoma. Finally, a potential treatment using adult stem cells, specifically dental stem cells, has highlighted the possibility of regenerating the damaged cornea frequently resulting from ASD. Future steps include further investigation of the AP-2 β NCC KO mouse mutants and determination of whether knockouts at different time points during embryonic development yield different clinical manifestations. Regenerative medicine, such as the use of DSCs, should be further validated and advanced to human trials in order to treat the millions affected by ASD.

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- Chavarría-Soley G, Michels-Rautenstrauss K, Caliebe A, Kautza M, Mardin C, Rautenstrauss B. Novel CYP1B1 and known PAX6 mutations in anterior segment dysgenesis (ASD). *Journal of glaucoma*. 2006 Dec 1;15(6):499-504.
- Liu P, Johnson RL. Lmx1b is required for murine trabecular meshwork formation and for maintenance of corneal transparency. *Developmental Dynamics*. 2010 Aug 1;239(8):2161-71.
- Pressman CL, Chen H, Johnson RL. LMX1B, a LIM homeodomain class transcription factor, is necessary for normal development of multiple tissues in the anterior segment of the murine eye. *Genesis*. 2000 Jan 1;26(1):15-25.
- Qiu Q, Chen H, Johnson RL. Lmx1b expressing cells in the mouse limb bud define a dorsal mesenchymal lineage compartment. *genesis*. 2009 Apr 1;47(4):224-33.
- Weng J, Luo J, Cheng X, Jin C, Zhou X, Qu J, Tu L, Ai D, Li D, Wang J, Martin JF. Deletion of G protein-coupled receptor 48 leads to ocular anterior segment dysgenesis (ASD) through down-regulation of Pitx2. *Proceedings of the National Academy of Sciences*. 2008 Apr 22;105(16):6081-6.
- Luo J, Zhou W, Zhou X, Li D, Weng J, Yi Z, Cho SG, Li C, Yi T, Wu X, Li XY. Regulation of bone formation and remodeling by G-protein-coupled receptor 48. *Development*. 2009 Aug 15;136(16):2747-56.
- Martino VB, Sabljic T, Deschamps P, Green RM, Akula M, Peacock E, Ball A, Williams T, West-Mays JA. Conditional deletion of AP-2 in mouse cranial neural crest results in anterior segment dysgenesis and early-onset glaucoma. *Disease Models & Mechanisms*. 2016 Aug 1;9(8):849-61.
- Chen L, Martino V, Dombkowski A, Williams T, West-Mays J, Gage P, AP-2 Is a Downstream Effector of PITX2 Required to Specify Endothelium and Establish Angiogenic Privilege During Corneal Development. *Investigative Ophthalmology & Visual Science*. 2016;57(3):1072.
- Evans AL, Gage PJ. Expression of the homeobox gene Pitx2 in neural crest is required for optic stalk and ocular anterior segment development. *Human molecular genetics*. 2005 Nov 15;14(22):3347-59.
- Cox CJ, Espinoza HM, McWilliams B, Chappell K, Morton L, Hjalil TA, Semina EV, Amendt BA. Differential regulation of gene expression by PITX2 isoforms. *Journal of Biological Chemistry*. 2002 Jul 12;277(28):25001-10.
- Amendt BA, Sutherland LB, Semina EV, Russo AF. The Molecular Basis of Rieger Syndrome ANALYSIS OF PITX2 HOMEODOMAIN PROTEIN ACTIVITIES. *Journal of Biological Chemistry*. 1998 Aug 7;273(32):20066-72.
- Suh H, Gage PJ, Drouin J, Camper SA. Pitx2 is required at multiple stages of pituitary organogenesis: pituitary primordium formation and cell specification. *Development*. 2002 Jan 15;129(2):329-37.
- Priston M, Kozlowski K, Gill D, Letwin K, Buys Y, Levin AV, Walter MA, Héon E. Functional analyses of two newly identified PITX2 mutants reveal a novel molecular mechanism for Axenfeld-Rieger syndrome. *Human molecular genetics*. 2001 Aug 1;10(16):1631-8.
- Sowden JC. Molecular and developmental mechanisms of anterior segment dysgenesis. *Eye*. 2007 Oct 1;21(10):1310-8.
- Oveki A, Wang WL. Retinoic acid signaling in mammalian eye development. *Experimental eye research*. 2009 Sep 30;89(3):280-91.
- Balkan W, Klintworth GK, Bock CB, Linney E. Transgenic mice expressing a constitutively active retinoic acid receptor in the lens exhibit ocular defects. *Developmental biology*. 1992 Jun 1;151(2):622-5.
- Balkan W, Klintworth GK, Bock CB, Linney E. Transgenic mice expressing a constitutively active retinoic acid receptor in the lens exhibit ocular defects. *Developmental biology*. 1992 Jun 1;151(2):622-5.
- Enwright JF, Grainger RM. Altered retinoid signaling in the heads of small eye mouse embryos. *Developmental biology*. 2000 May 1;221(1):10-22.
- Trainor P, editor. *Neural crest cells: Evolution, development and disease*. Academic Press; 2013 Nov 23.
- Kumar S, Duester G. Retinoic acid signaling in periopitic mesenchyme represses Wnt signaling via induction of Pitx2 and Dkk2. *Developmental biology*. 2010 Apr 1;340(1):67-74.
- Yam GH, Peh GS, Singhal S, Goh BT, Mehta JS. Dental stem cells: a future asset of ocular cell therapy. *Expert reviews in molecular medicine*. 2015;17:e20.
- Perry BC, Zhou D, Wu X, Yang FC, Byers MA, Chu TM, Hockema JJ, Woods EJ, Goebel WS. Collection, cryopreservation, and characterization of human dental pulp-derived mesenchymal stem cells for banking and clinical use. *Tissue Engineering Part C: Methods*. 2008 Jun 1;14(2):149-56.
- Syed-Picard FN, Du Y, Lathrop KL, Mann MM, Funderburg ML, Funderburg JL. Dental pulp stem cells: a new cellular resource for corneal stromal regeneration. *Stem cells translational medicine*. 2015 Mar;4(3):276-85.